Curcumin sensitizes prostate cancer cells to tumor necrosis factor–related apoptosis-inducing ligand/Apo2L by inhibiting nuclear factor-κB through suppression of IκBα phosphorylation

Dorrah Deeb,1 Hao Jiang,2 Xiaohua Gao,1 Mikehl S. Hafner,3 Henry Wong,3 George Divine,4 Robert A. Chapman,5 Scott A. Dulchavsky,1 and Subhash C. Gautam1

Departments of 1Surgery, 2Neurology, 3Dermatology, and 4Biostatistics and Research Epidemiology and 5Division of Hematology/Oncology, Henry Ford Health System, Detroit, Michigan

Abstract
Epidemiologic studies suggest that diet rich in plant-derived foods plays an important role in the prevention of prostate cancer. Curcumin, the yellow pigment in the spice turmeric, has been shown to exhibit chemopreventive and growth inhibitory activities against multiple tumor cell lines. We have shown previously that curcumin and tumor necrosis factor–related apoptosis-inducing ligand (TRAIL)/Apo2L interact to induce cytotoxicity in the LNCaP prostate cancer cell line. In this study, we investigated the mechanism by which curcumin augments TRAIL-induced cytotoxicity in LNCaP cells. Subtoxic concentrations of the curcumin-TRAIL combination induced strong apoptotic response in LNCaP cells as demonstrated by the binding of Annexin V-FITC and cleavage of procaspase-3. Furthermore, LNCaP cells express constitutively active nuclear factor-κB (NF-κB), which is inhibited by curcumin. Because NF-κB has been shown to mediate resistance to TRAIL-induced apoptosis in tumor cells, we investigated whether there is a relationship between NF-κB activation and resistance to TRAIL in LNCaP prostate cancer cells. Pretreatment with curcumin inhibited the activation of NF-κB and sensitized LNCaP cells to TRAIL. A similar increase in the sensitivity of LNCaP cells to TRAIL-induced apoptosis was observed following inhibition of NF-κB by dominant negative mutant IκBα, an inhibitor of NF-κB. Finally, curcumin was found to inhibit NF-κB by blocking phosphorylation of IκBα. We conclude that NF-κB mediates resistance of LNCaP cells to TRAIL and that curcumin enhances the sensitivity of these tumor cells to TRAIL by inhibiting NF-κB activation by blocking phosphorylation of IκBα and its degradation. [Mol Cancer Ther 2004;3(7):803–12]

Introduction
Prostate cancer is the second leading cause of cancer-related deaths in the United States (1, 2). Androgen reduction therapy is largely effective against hormone-sensitive tumors, but hormone-refractory clones often emerge after hormone therapy (3). Because apoptosis is the underlying mechanism by which most anticancer therapies including chemotherapy, radiation, and antihormonal therapy kill tumor cells, novel agents that may sensitize drug-resistant tumor cells for induction of apoptosis by conventional therapies could lead to the regression and improved prognosis of the refractory disease (4, 5). Indeed, chemotherapeutic agents have been shown to sensitize tumor cells to killing by death ligands such as tumor necrosis factor-α and Fas ligand (ApotI or CD95L; refs. 6, 7). However, the use of these apoptosis-inducing antineoplastic agents is limited by unacceptable systemic toxicity. Tumor necrosis factor–related apoptosis-inducing ligand (TRAIL)/ApotI2L, a recently discovered member of the tumor necrosis factor superfamily, is a potent inducer of apoptosis in various cancer cell lines including prostate, colon, breast, skin, kidney, lung, brain, and hematologic tumor cell lines, with little cytotoxicity to the normal cells (8, 9). TRAIL is also effective in inhibiting the tumor growth in vivo (10, 11), and unlike the severe inflammatory response syndrome induced by tumor necrosis factor-α and the hepatotoxicity of Fas ligand (12, 13), treatment of mice and nonhuman primates with TRAIL is without systemic toxicity (14). Recently, however, recombinant human TRAIL was shown to induce massive apoptosis in normal human liver hepatocytes but not in hepatocytes isolated from the livers of mice, rat, or nonhuman primates (15). The hepatotoxicity of soluble recombinant human TRAIL used in this study, as well as its toxicity toward normal keratinocytes shown in another report (16), was shown to result from the modifications introduced to the TRAIL molecule to stabilize and enhance trimerization of the three subunits of TRAIL. It is clear now that only the polyhistidine tagged or leucine zipper motif containing versions of recombinant human TRAIL with low zinc contents are toxic to the normal cells (17, 18).
Although the exact cause of prostate cancer is not known, epidemiologic studies suggest that, in addition to race and age, diet is a prominent risk factor for prostate cancer (19). While a high-fat diet has been implicated in the occurrence of human cancers, including prostate cancer, several polyphenolic compounds found in common plant-derived foods have been recognized for their anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric extracted from the rhizome of the plant *Curcuma longa*, has been shown to have strong anti-inflammatory, antioxidant, and anti-lipooxygenase/cyclooxygenase activities (21, 22). Because of its ability to scavenge free radicals and inhibit inflammation, curcumin has been investigated for chemotherapeutic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20).

Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20).

Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20). Curcumin (diferuloyl-methane), the yellow pigment found in the spice turmeric, has been recognized for its anticarcinogenic properties (20)
(anti–CD16/CD32), and reacted with primary goat anti-TRAIL-R1, anti-TRAIL-R2, anti-TRAIL-R3, or anti-TRAIL-R4 antibodies (5 μg/mL) for 45 minutes on ice. Cells were washed once with PBS and incubated with phycoerythrin-conjugated donkey anti-goat secondary antibody (1:50 dilution) for 45 minutes at 4°C. Cells were washed and analyzed by flow cytometry.

Isolation of Nuclear Proteins

Nuclear extracts were prepared by the modified procedure of Dignam et al. (42). Following treatment with curcumin for 24 hours, LNCaP cells were washed three times with PBS and incubated on ice for 15 minutes in hypotonic buffer A [10 mmol/L HEPES (pH 7.9), 10 mmol/L KCl, 0.1 mmol/L EDTA, 0.1 mmol/L EGTA, 1 mmol/L DTT, 0.5 mmol/L phenylmethylsulfonyl fluoride, and 0.6% NP40]. Cells were vortexed gently for lysis, and nuclei were separated from the cytosol by centrifugation at 12,000 × g for 1 minute. Nuclei were resuspended in buffer C [20 mmol/L HEPES (pH 7.9), 25% glycerol, 0.4 mol/L NaCl, 1 mmol/L EDTA, 1 mmol/L EGTA, 1 mmol/L DTT, and 0.5 mmol/L phenylmethylsulfonyl fluoride] and shaken for 30 minutes at 4°C. Nuclear extracts were obtained by centrifugation at 12,000 × g, and protein concentration was measured by Bradford assay (Bio-Rad, Richmond, CA). NF-κB in nuclear extracts was detected by Western blotting as described below.

NF-κB Binding Activity

NF-κB (p65) binding activity in nuclear extracts was measured using the ELISA-based TransAm NF-κB kit. In this assay, NF-κB binds to the immobilized oligonucleotide containing the NF-κB consensus sequence (5′-GGGA-CTTTCC-3′), which is detected by sandwich ELISA. The detection limit for TransAm NF-κB kit is <0.4 ng/mL purified p65.

Western Blot Analysis

Total cellular proteins were isolated by detergent lysis [1% Triton X-100 (v/v), 10 mmol/L Tris-HCl (pH 7.5), 5 mmol/L EDTA, 150 mmol/L NaCl, 10% glycerol, 2 mmol/L sodium vanadate, 5 μg/mL leupeptin, 1 μg/mL aprotinin, 1 μg/mL pepstatin, and 10 μg/mL 4-2-aminoethyl-benzenesulfonyl fluoride] as described by Keane et al. (43). Lysates were clarified by centrifugation at 14,000 × g for 10 minutes at 4°C, and protein concentrations were determined by Bradford assay. Samples (50 μg) were boiled in an equal volume of sample buffer [20% glycerol, 4% SDS, 0.2% bromophenol blue, 125 mmol/L Tris-HCl (pH 7.5), and 640 mmol/L 2-mercaptoethanol] and separated on 10% to 14% SDS-PAGE gels. Proteins resolved on the gels were transferred to polyvinylidene difluoride membranes. Membranes were blocked with 5% milk in 10 mmol/L Tris-HCl (pH 8.0) and 150 mmol/L NaCl with 0.05% Tween 20 and probed with protein specific antibodies to caspase-3, poly(ADP-ribose) polymerase, NF-κB (p65), or phospho-IκBα. Immunocomplexes were visualized with enhanced chemiluminescence detection system from Amersham (Arlington Heights, IL).

Intracellular Localization of NF-κB

LNCaP cells cultured on cover slips for 24 hours were treated with curcumin (50 μmol/L) for 4 hours. After treatment, cells were washed twice and fixed with cold 4% paraformaldehyde for 1 hour. After washing in PBS, cells were blocked, permeabilized, and incubated overnight at 4°C with rabbit anti-RelA (p65) antibody (1:500 dilution). After incubation, cells were treated with Texas red–conjugated anti-rabbit IgG in the dark at room temperature for 2 hours. Cells were washed, mounted with mounting medium containing 4′,6-diamidino-2-phenylindole, and examined under fluorescent microscope.

DNA Transfections

The transcripational activity of constitutive nuclear NF-κB was measured in NF-κB-dependent luciferase reporter gene expression assay. Briefly, LNCaP cells were plated in six-well plates for 24 hours and transiently transfected using the LipofectAMINE reagent (Invitrogen) with pκB-Luc (2 μg DNA) expression plasmid containing three HIV κB sites upstream of the thymidine kinase minimal reporter and the luciferase cDNA. Cells were incubated at 37°C for 24 hours and lysed to prepare cell extracts. The luciferase activity in cell extracts (50 μL) was measured in a luminometer using reagents and instructions provided with the luciferase assay system (Promega). In some cultures, LNCaP cells were also transfected with pCMV-κBα vector (2 μg) expressing the dominant negative mutant IκBα (A32/36) from Clonetech (Palo Alto, CA). The mutant IκBα contains serine-to-alanine mutations at residues 32 and 36, which confers resistant to phosphorylation and subsequent proteosome-mediated degradation but still binds to NF-κB.

Statistical Analysis

Data are presented as means ± SD. Interaction between TRAIL and curcumin was tested by two-way ANOVA. The degree of interaction was expressed as the percentage difference between the combined TRAIL and curcumin response and the sum of the responses to TRAIL and curcumin alone.

Results

Combined Curcumin and TRAIL Treatment Induces Cytotoxicity in Prostate Cancer Cells

The induction of cytotoxicity in LNCaP prostate cancer cells by curcumin (10 and 25 μmol/L), TRAIL (20 ng/mL), and combination of curcumin and TRAIL was calculated from the loss of cell viability measured by the MTS assay. As shown in Fig. 1A, treatment of LNCaP cells with curcumin (10 or 25 μmol/L) or TRAIL alone induced little cytotoxicity (~10%). In contrast, simultaneous exposure of LNCaP cells to curcumin at both 10 and 25 μmol/L and TRAIL resulted in significantly enhanced cytotoxicity (52% and 77%, respectively). On the other hand, the viability of PZ-HPV-7 human prostatic epithelial cells, which are transformed but are not tumorigenic, was not affected at these concentrations of curcumin or TRAIL when applied separately or in combination (data not shown). Overall, the
amount of cytotoxicity induced with combined curcumin and TRAIL treatment was 320% and 489% higher for 10 and 25 μmol/L curcumin-TRAIL combinations than cytotoxicity that would be induced if the combined effect was simply additive.

To determine whether apoptosis is the mode of cell death induced by combination of curcumin and TRAIL, LNCaP cells treated with curcumin, TRAIL, or curcumin-TRAIL combination for 48 hours were examined by flow cytometry for binding of Annexin V-FITC to phosphatidylserine, which is externalized to the outer surface of the plasma membrane in cells undergoing apoptosis. As shown in Fig. 1B, there was little binding of Annexin V-FITC to untreated LNCaP cells or those treated with curcumin alone (9% and 12%, respectively). The binding of Annexin V-FITC was increased somewhat following treatment of cells with TRAIL (21%). However, the binding of Annexin V-FITC with combined treatment with curcumin and TRAIL (58%) was 76% higher than what would be expected if the effect was merely additive. The P value for the test for interaction was 0.003.

To further investigate induction of apoptosis, we performed immunoblot analysis of whole cell lysate to determine cleavage of procaspase-3. Untreated LNCaP cells and those treated with curcumin (10 μmol/L) for 20 hours showed little or no cleaved fragments (19 and 17 kDa) of caspase-3 (Fig. 1C, lanes 1 and 2). The cleavage of procaspase-3 was increased in cells treated with TRAIL alone (20 ng/mL; lane 3). However, by far the most processing of procaspase-3 was observed in cells treated with curcumin-TRAIL combination (lane 4). Thus, enhanced binding of Annexin V-FITC and activation of effector caspase-3 demonstrate induction of apoptosis in cells treated with curcumin-TRAIL combination.

**Effect of Curcumin on Expression of TRAIL-R on LNCaP Cells**

Because TRAIL-induced apoptosis in LNCaP cells was enhanced by curcumin, we considered the possibility that curcumin might enhance apoptosis by TRAIL by modulating the expression TRAIL-R. TRAIL transmits the proapoptotic signal by interacting with death receptor 4 (DR4 or TRAIL-R1) and death receptor 5 (DR5 or TRAIL-R2). Both TRAIL-R1 and TRAIL-R2 have cytoplasmic death domains that can transduce death signals and activate caspases and NF-κB (44). TRAIL also binds to decoy receptors DcR1 (TRAIL-R3) and DcR2 (TRAIL-R4), which have truncated death domains and are incapable of transducing death signals (45). We examined LNCaP cells for the expression of TRAIL-R and the effect curcumin has on their expression by flow cytometry. Figure 2 shows that LNCaP cells express all of the four TRAIL-R. The high-affinity death signaling TRAIL-R2 is more abundantly expressed than TRAIL-R1. In addition, both decoy receptors (TRAIL-R3 and TRAIL-R4) are also heavily expressed on LNCaP cells. Treatment with curcumin (10 μmol/L) for 48 hours up-regulated the expression of TRAIL-R without altering the expression of TRAIL-R2. Curcumin also increased the expression of TRAIL-R3 without affecting TRAIL-R4 expression. These results indicate that curcumin selectively increases the expression of TRAIL-R1 and TRAIL-R3 on LNCaP cells.

**LNCaP Cells Express NF-κB and Curcumin Inhibits It**

We next investigated the possibility that constitutively active NF-κB mediates the resistance of LNCaP cells to
TRAIL, and potentiation of TRAIL killing of these cells might involve inhibition of NF-κB activation by curcumin. Immunoblot analysis of nuclear extracts prepared from untreated and treated cells revealed the expression of constitutive NF-κB, which was inhibited following treatment with curcumin for 20 hours in a dose-dependent manner (Fig. 3A). NF-κB was also suppressed in cells treated with curcumin-TRAIL combination. On the other hand, treatment with TRAIL alone had no effect on NF-κB expression. Similar results were obtained in experiments in which DNA binding activity of NF-κB to the κB consensus sequence was measured. These measurements also showed activated NF-κB in LNCaP cells, which is inhibited by curcumin (Fig. 3B). These results were further corroborated by immunocytochemical examination of NF-κB in LNCaP cells. Control cells exhibited intense nuclear localization of NF-κB as detected by treatment with anti-RelA (p65) primary antibody followed by staining with Texas red–conjugated secondary antibody (Fig. 3C). Cells treated with curcumin (50 μmol/L) for 4 hours showed diffused cytoplasmic expression of RelA. Taken together, these results demonstrate that LNCaP cells express constitutively active NF-κB, which is inhibited by curcumin.

Constitutive NF-κB in LNCaP Cells Is Transcriptionally Active

To examine whether constitutive NF-κB is transcriptionally active, LNCaP cells were transiently transfected with NF-κB regulated p3κB-Luc reporter construct to measure NF-κB-dependent reporter gene expression. As shown in Fig. 4, the relative luciferase activity of LNCaP cells transfected with p3κB-Luc reporter vector was ~75-fold higher compared with cells transfected with the control plasmid without κB sites. The luciferase activity was suppressed when transfected cells were treated with curcumin at 50 μmol/L. Luciferase activity was also suppressed when cells were double transfected with p3κB-Luc and dominant negative pCMV-IκBαM (mutant) vectors. The inhibition of NF-κB activity by curcumin or pCMV-IκBαM was further confirmed by immunoblotting of nuclear NF-κB (top). These results indicate that constitutive NF-κB in LNCaP cells is transcriptionally active.

Pretreatment with Curcumin Sensitizes LNCaP Cells to TRAIL by Inhibiting NF-κB

To determine whether the mechanism by which curcumin sensitizes LNCaP cells to TRAIL-induced apoptosis involves inhibition of NF-κB, we pretreated LNCaP cells with curcumin at 25 μmol/L for 48 hours followed by extensive washing. Cells were treated at 25 μmol/L curcumin because, at this concentration, curcumin almost completely blocks NF-κB expression (Fig. 3A). After treatment with curcumin, cells were incubated with TRAIL (20 ng/mL) for 48 hours to determine cytotoxicity. As
shown in Fig. 5A, pretreatment with curcumin for 48 hours resulted in the suppression of NF-κB (day 0). Following the removal of curcumin from cultures, NF-κB began to recover and reached the pretreatment levels between days 6 and 10. The induction of sensitivity to TRAIL in pretreated cells was tightly associated with the level of NF-κB (Fig. 5B). Induction of cytotoxicity in pretreated cells was most pronounced on day 0 (58%), a time at which NF-κB was maximally suppressed. The progressive decrease in percentage cytotoxicity from days 2 to 10 corresponded with the reappearance of nuclear NF-κB in treated cells. Thus, these data demonstrate a clear correlation between constitutive NF-κB and resistance/sensitivity to TRAIL.

Inhibition of NF-κB by Ectopic Expression of IκBα Restores Susceptibility of LNCaP Cells to TRAIL

Results shown in Fig. 5 demonstrated that the level of NF-κB determines the resistance or susceptibility of LNCaP cells to TRAIL. These data, however, do not provide direct evidence that NF-κB mediates resistance of LNCaP cells to TRAIL because, besides inhibiting NF-κB, curcumin may also induce additional biochemical changes in cells that could contribute to the sensitization of LNCaP cells to TRAIL. To test this more directly, LNCaP cells were transiently transfected with pCMV-IκBα expression vector (2 µg plasmid DNA) for 20 hours. Cells were extensively washed in PBS to remove curcumin and tested for expression of nuclear NF-κB and susceptibility to TRAIL. The expression of NF-κB was dramatically reduced on day 2 after transfection and returned to the pretreatment level by day 12 (Fig. 6A). Day 2 cells with highly decreased level of NF-κB showed significantly enhanced susceptibility to TRAIL in the absence of curcumin (Fig. 6B). On the other hand, day 12 transfected cells expressed NF-κB at a level comparable with that for control cells and were resistant to TRAIL. However, these cells remained susceptible to the combined curcumin-TRAIL treatment. These results provide direct evidence that NF-κB mediates susceptibility of LNCaP cells to TRAIL.

Curcumin Inhibits IκBα Phosphorylation

To determine whether inhibition of NF-κB by curcumin is due to inhibition of the degradation of IκBα, we measured the effect of curcumin on phosphorylation of IκBα, an essential event for its proteolytic degradation. As shown in Fig. 7A, treatment of LNCaP cells with TRAIL increased the phosphorylation of IκBα (lane 2 versus lane 1). Phosphorylation of IκBα was partially to completely inhibited by curcumin at 12.5 µmol/L (lane 2) after 4- and

---

**Figure 4.** Constitutive NF-κB is transcriptionally active in LNCaP cells. LNCaP cells grown to 60% to 70% confluency were transfected with control vector (pGL3-Luc), p3xκB-Luc, or pκBα-mut and dominant negative IκBαM (κBα-mut) expression vectors (2 µg plasmid DNA) for 24 hours. Culture medium was replaced with fresh medium, and in some wells, cells transfected with p3xκB-Luc vector were treated with curcumin (50 µmol/L) for 20 hours. Cell lysates were prepared, and luciferase activity was determined in a luminometer using the luciferase assay system. Nuclear extracts prepared from p3xκB-Luc transfected LNCaP cells treated with curcumin or transfected with IκBα expression vector were analyzed for NF-κB by Western blotting (top). Similar results were obtained in two separate experiments.
20-hour treatment, respectively. At higher concentrations (25 to 50 μmol/L), curcumin completely inhibited IκBα phosphorylation (lanes 4 and 5) after 4-hour treatment. These data imply that curcumin inhibits constitutive NF-κB by suppressing phosphorylation and degradation of IκBα.

Discussion
Recent studies have shown that plant-derived dietary compounds provide protection against the development of cancers and other diseases (46, 47). We and others have shown previously that food additive curcumin inhibits proliferation and induces apoptosis in several tumor cell lines (26-30). We have also demonstrated that curcumin sensitizes hormone-sensitive LNCaP human prostate cancer cells to TRAIL-induced apoptosis by promoting cleavage of procaspases and release of cytochrome c from mitochondria (32). Here, we report that suppression of the constitutively active NF-κB in LNCaP cells is a major event by which curcumin chemosensitizes prostate cancer cells to TRAIL-induced cytotoxicity. The blockade of NF-κB activation by curcumin involved repression of the phosphorylation of IκBα, an event that primes IκBα for proteasomal degradation.

Figure 6. Expression of dominant negative IκBα inhibits NF-κB and increases susceptibility to TRAIL. LNCaP cells were transfected with dominant negative pCMV-IκBα vector expressing mutant IκBα (A32/36). Transfected cells were harvested to measure NF-κB and sensitivity to TRAIL. A, western blot analysis of NF-κB in control and transfected cells (days 2 and 12). B, cytotoxic response of days 2, 6, and 12 transfected cells to TRAIL (20 ng/mL) or day 12 cells to curcumin-TRAIL combination (cur/TRAIL; hatched bars) and cytotoxic response of control LNCaP cells to TRAIL, curcumin, or combined curcumin-TRAIL (solid bars). Similar results were obtained in three independent experiments.

Figure 7. Curcumin inhibits phosphorylation of IκBα. A, LNCaP cells were treated with TRAIL (20 ng/mL) or combined TRAIL-curcumin (12.5, 25, and 50 μmol/L) simultaneously for 4 or 20 hours. Cell lysates were prepared and analyzed by Western blotting to determine phosphorylated IκBα using antibodies against phosphorylated IκBα. B, histogram showing the relative levels of phospho-IκBα. Identical results were obtained in two separate experiments.

Mol Cancer Ther 2004;3(7). July 2004
sensitivity to TRAIL because inhibitory TRAIL-R3 was also increased by curcumin. It should be noted, however, that TRAIL-R data presented are the percentage of cells expressing each receptor, not the total receptor abundance. Therefore, the possibility remains that curcumin sensitizes LNCaP cells to TRAIL by altering the balance of death and decoy TRAIL-R in favor of death-inducing TRAIL-R1 and TRAIL-R2. NF-κB not only plays a critical role in the transcription of genes involved in immune and inflammatory responses (33), cell proliferation/differentiation (53), or cell transformation (54) but also acts as a survival factor by protecting tumor cells by inhibiting apoptosis (34-37). Because NF-κB protects tumor cells from apoptosis, we considered the possibility that resistance of prostate cancer cells to TRAIL might also be due to increased NF-κB activity. LNCaP cells were found to express transcriptionally active NF-κB. Treatment with curcumin inhibited NF-κB in a dose-dependent fashion. Others have also shown curcumin to suppress constitutive and inducible NF-κB activity in several cell types (54-56). Because activated NF-κB induces proliferation in tumor cells and therefore curtails apoptosis, this might explain why LNCaP cells are resistant to TRAIL.

Our results also demonstrate that constitutively active NF-κB in LNCaP cells is not merely an epiphenomenon irrelevant to the resistance of these cells to TRAIL, because suppression of NF-κB through pretreatment with curcumin or transfection with superrepressor IkBα (A32/36) sensitized LNCaP cells to TRAIL. This result agrees with previous studies showing that constitutive NF-κB renders tumor cells resistant to TRAIL (54, 57, 58), and its inhibition with various agents leads to the sensitization of tumor cells to TRAIL-induced apoptosis (59-61).

The mechanism by which curcumin inhibits NF-κB is unclear. In resting cells, NF-κB remains sequestered in the cytoplasm in a functionally inactive form noncovalently bound to an inhibitory protein, IκBα (62). On stimulation of cells with mitogens, antigens, or cytokines, IκBα is phosphorylated and degraded allowing NF-κB to translocate to the nucleus where it binds to the κB motifs in the promoter region of the responsive genes. Our result demonstrating the inhibition of phosphorylation of IκBα indicates that curcumin inhibits NF-κB signaling by preventing IκBα phosphorylation and its degradation. These results are consistent with previous reports in which curcumin was shown to inhibit phosphorylation of IκBα (54, 63). Whether curcumin inhibits IκBα phosphorylation in LNCaP cells by blocking IKK activity, the kinase that phosphorylates IκBα remains to be determined.

It is also not entirely clear how suppression of NF-κB by curcumin promotes apoptosis, but it may be related to the effect of curcumin on NF-κB target genes that regulate apoptosis. Curcumin down-regulates the expression of NF-κB-dependent antiapoptotic Bcl-2 and Bcl-xl in prostate cancer cell lines (29). The inhibition of NF-κB by curcumin may also attenuate members of the inhibitor of apoptosis proteins family, which are regulated by NF-κB (64, 65). Besides NF-κB, other molecular targets of curcumin such as cell survival signal protein kinase B/Akt, c-Jun NH2-terminal kinase, and peroxisome proliferator-activated receptor γ (66-68) may also contribute to the chemosensitization of prostate cancer cells to TRAIL.

In conclusion, present studies demonstrate that constitutively active NF-κB mediates resistance of prostate cancer cells to TRAIL, and the mechanism by which curcumin sensitizes them to TRAIL involves suppression of NF-κB activation through inhibition of IκBα phosphorylation and its degradation. Although the finding that a relatively nontoxic agent such as curcumin can restore the responsivity of LNCaP cells to TRAIL is promising, additional prostate cancer cell lines will have to be tested before the combined curcumin-TRAIL treatment regimen could be considered for clinical application. It is also unclear whether the effective (sensitizing) concentration of curcumin can be achieved in vivo because of the poor bioavailability of orally administered curcumin (69). This difficulty may be surmounted, however, by coformulation with piperine, which has been shown to inhibit hepatic and intestinal glucuronidation and increase the bioavailability of curcumin (70).

Acknowledgments
We thank Carolyn Hedgcock for assistance with luciferase reporter gene assay.

References


44. Schneider P, Thome M, Burns K, et al. TRAIL receptors 1 (DR4) and 2 (DR5) signal FADD-dependent apoptosis and activate NF-κB. Immunity 1997;7:831-6.


